Toxicology of Hydraulic Fracturing Flowback Fluid - Potential Endocrine Disruptor?





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Hydraulic Fracturing Process

- Mixture of water, proppants and chemical additives are injected into the ground at high pressures (~70,000 kPa)
- Proppants prevent the fractures from closing
- Chemical additives include biocides and surfactants

Shale

 When well is opened, 10-70% of injected water returns to surface

~ 0.5 -4 km

Flowback and Produced Waters

Distinction between terms Flowback and Produced Water (FPW) are operational only. FPW varies by:

- 1. Time of flowback
 - earlier has more anthropogenic derived chemicals
 - later have more formation water
- 2. Formation characteristics
 - inorganic and petrogenic components
- 3. Potential "downhole" reaction products
 - heat, chemicals and pressure

Chemical compositions of injecting mixtures are commonly held as trade secrets by HF practitioners

Duvernay Region- Alberta-

- Anywhere between 10 -100 million litres of fracturing fluid and water pumped down into each well
 10 20% of injected fluid comes back up as flowback/produced
- 10 80% of injected fluid comes back up as flowback/produced water



Distribution map for cumulative injected water (in m³) per well, for wells fractured between November 2011 and March 2014

Canadian Water Network- Goss et al 2015, Alessi et al 2016 Can Water Resour J



Numerous large and small FPW spills into freshwater lakes or wetlands have been documented

Zama City, Alberta, 2013, ~9.5 million litres of contaminated FPW





Natural Sciences and Engineering Research Council of Canada Collaborative Research and Development Program Three co-PIs: Daniel Alessi, Jon Martin and Greg Goss Industrial partner: Encana Corp

Aims:

- 1. Comprehensive profiling of FPW chemistry and toxicology
- 2. Comprehensive characterizations and assessment of the geological formation and production of new chemical species
- 3. Provide feedback to Encana about toxicity of FPW (temporal and geological)



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Flowback and Produced Water samples

Samples provided by Encana

AC- Activated Carbon treatment (removes organic and most metals)

SF- sediment free Filtered 0.2 μM

FPW - from Paskapoo formation (AB) significant sediment





Chemical characterization of FPW-inorganics

 Inorganic parameters measured by multiple methods (Alessi Lab)

Note: salinity of 242,000 mg/L is ~7 X seawater (~33,000 mg/L)

7-day FPW Sample (pH = 4.78)		
Parameters	Method	Mean Conc. (mg/L)
TDS	Evaporation	242,624
TN	TC/TN Analyzer	498
тс	TC/TN Analyzer	211
Na	ICP-MS/MS	70,000
Са	ICP-MS/MS	11,800
К	ICP-MS/MS	2,570
Cl	IC	136,000



Sediment Fraction in FPW contained hollow microspheres



SEM image of spherical structure (1) and presumed fragments (2) with EDX elemental analysis of exterior of sphere, respectively.

Exterior

Carbon 64% Oxygen 33% Iron 2% Silica 1% Calcium <1%.

Thought to contain a "breaker" such as ammonium persulfate (oxidant) ** disclaimer**

Close up SEM image of a broken sphere showing its hollow nature and thin shell walls (3) with EDX elemental analysis of the interior wall of a sphere.

Interior: Carbon 79.5%, Oxygen 19%, iron and silica <1%



Organic Analysis (Untargeted - Orbitrap)



Possible formation of downhole reaction products

Downhole conditions





Irgaphos 168 Tris(2,4-di-tert-butylphenyl) phosphiteantioxidant chemical stabilizer

Note: we do not know if this was the chemical in the original HF fluids or if the phosphates were added directly. Detected the above organophosph<u>ate</u> chemicals in our FPW sample, (believe it is *via* oxidation of the parent phosphite compound) NB: still needs investigation

₄H_o

1

HO



 C_4H_2

 C_4H

Toxicological assessment



Rainbow Trout (Oncorhynchus mykiss) (

Zebrafish embryo (*Danio rerio*) model Water flea (*Daphnia magna*)

Common Ecotoxicological models (Environment Canada, OECD, US EPA)

- Provide information re. ecological hazard and risk assessment
- Zebrafish embryo exposure model used to assess possible early life-stage effects



Methods







Rainbow trout fingerlings

- 24 or 48 hour acute exposure
- 0 (control), 2.5% or 7.5% FPW
- tested both sediment containing and sediment free (filtered) FPW



Biotransformation enzymes



Ethoxyresorufin-o-deethylase (EROD) a phase 1 biotransformation enzyme used to eliminate planar hydrocarbons

- common marker of exposure to oil and gas contaminated waters
- BaP exposure (right bar) simply used to demonstrate our assay was working properly
- Induction in gill greater than in liver
- Induction in gills higher with sediment containing FPW compared to sediment free



Oxidative stress (TBARS formation)



- The FPW alone can generate free oxygen radicals
- Greater at 7.5% compared to 2.5%
- No difference with or without sediment
- All tissues (liver, gill, kidney) showed evidence of exposure to greater free oxygen radicals (through TBARS formation in lipids)
- Greater response at 7.5% compared to 2.5%



Endocrine disruption



Gene responses in liver of rainbow trout exposed to sediment containing 2.5% and 7.5% FPW

Note large increased in expression of egg yolk protein (Vtg) and Estrogen receptor a2 (era2) but not estrogen receptors era), erb1 or erb2

Acute Toxicity analysis - zebrafish embryo

- FPWs pose strong acute toxicity to ZF embryo
 - During 1-24h, lethality LC50 FPW 0.6%, SF 0.92% was driven by organic contents
 - After 24h, acute toxicity is dominated by salt
- Sediment containing had great toxicity compared to sediment free solutions
- Toxicological profile greatly complicated by salinity of FPW





ZF embryo after 24h exposure (2.5%)



Methods



Zebrafish embryo (*Danio rerio*) model

Objectives

- Compare new samples of FPW received from two different well for toxicological profile and underlying possible effects
- Assess toxicological profile without salinity induced effects
 - further examine the differences between sediment-containing and sediment-free FPW

Solution - separate and extract organics from each sample **Caveat**: we recognize that filtration of FPW under vacuum likely removes significant amounts of lower molecular weight volatile organics

Experimental design: Extraction of Organic Fractions of HF-FPW



Results: Total PAHs in Organic Fractions (ng/L)





Acute Toxicity to ZF embryo (168 hpf)



General Toxicity

- #2 FPW > #1
 FPW
- P > W > S

	LC50 (%)
1S	2630%
1W	1450%
1P	1200%
25	720%
2W	480%
2P	280%



CCS = 2 x 3 + 3 x 4 = 18





- $2P > 2W \ge 2S$
- $1P > 1W \ge 1S$
- #2 > #1



Transcriptional Response – Cytochrome p450s



Transcriptional Response - Oxidative stress



 Induction of oxidative stress responsive gene is consistent with the morphological observation of pericardial edema

(gstp1)

1S

1P

2W 2S

2P

25

8

Transcriptional Response – Endocrine disruption





Daphnia magna (water flea)

Daphnia are a highly sensitive FW species to FPW LC₅₀ of FPW of <1%

Neonates more sensitive that adults

Paired salt water controls (SW and AC) still have significant toxicity.



Acute LC₅₀ Results



Effects on Daphnia magna reproduction



Chronic 30-day daphnia reproduction tests showed salinity alone affected reproduction. However, FPW further decreased reproduction, even at 0.004%



Effects of FPW on Daphnia gene expression



Lack of Cyp4 induction suggests minimal planar hydrocarbon exposure Increased expression of Cut gene – involved in molting and reproduction

Summary



Our early stage results suggest significant and multiple effects of FPW on three model aquatic species.

- We demonstrate *apparent* transformation products in FPW.
- We show that the sediment fraction of these FPW samples has greater toxicity than the water-accomodated fraction.



HF-FPW exposure effects may include: induction of biotransformation enzymes oxidation of lipids and metabolic impairment endocrine disruption

We cannot yet attribute any of these effects to a specific HF component.

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Research Papers

- *He Y*, *Folkerts EJ*, Zhang Y, Martin JW, Alessi DS, Goss GG. (2017). Effects on Biotransformation, Oxidative Stress, and Endocrine Disruption in Rainbow Trout (Oncorhynchus mykiss) Exposed to Hydraulic Fracturing Flowback and Produced Water. Submitted to *Environ Sci Technol* 51(2): 940-947 (doi: 10.1021/acs.est.6b04695).
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